

## Taq DNA Polymerase with Robust Buffer (+ dNTPs)

Product code	02-002 200 U 02-002-5 200	U x 5	
Size	200 U		
Storage	-20°C		
Concentration	5 U/µl		
Product	Thermus aquaticus DNA polymerase (Taq DNA polymerase) was expressed in E. coli in large		
Description	quantities and highly purified. The enzyme has thermostable DNA polymerase activity and the MW is 94 kDa. This enzyme is suitable for PCR reactions; capable of amplifying DNA		
	with various primers.		
Definition of	One unit is defined as the amount of enzyme that can incorporate 10nmols of total dNTPs		
activity	into an acid-insoluble material in 30 minutes at 74°C when activated salmon sperm DNA was		
•	used as template / primer.		
Purity	Greater than 95% purity as determined by SDS-PAGE (CBB staining) (Fig.1)		
	The absence of endonucleases and e	xonucleases was confirmed.	
PCR Test	Good amplification result was obtained in PCR reaction using $\lambda DNA$ as a template up to 14		
	kB (Fig.2)		
Components	Taq DNA polymerse (5U/µl): 20 mM Tris-HCl (pH 8.0), 100 mM KCl, 0.1 mM EDTA, 1 mM		
	DTT, 50% glycerol, 0.5% Tween20, 0.5% Igepal CA-630 (02-Taq 40µl)		
	10 x Robust Buffer (Taq) (02-Trb 1.0ml)		
	<b>2.5 mM (each) dNTPs</b> (02-Dnt 640 µl)		
Application	1. High-throughput PCR	General composition of PCR reaction mixture (total 50µl)	
	2. Colony PCR	Taq DNA polymerase (5 U/µl) (02-Taq)	
	3. Incorporation of dUTP, dITP,	$10x$ Robust Buffer (Taq) (02-Trb) $5 \ \mu l$ $2.5 \text{mM}$ (each) dNTPs (02-Dnt) $4 \ \mu l$	
	and fluorescence-labeled	Template <500 ng Primer 1 0.2~1.0 mM (final conc.)	
	nucleotides	Primer 2 0.2~1.0 mM (final conc.)	
	4. Primer extension	Sterile distilled water up to 50 µl	
	5. Addition of a single nucleotide (adenosine) at the 3'-blunt ends	X Use of excess amount is not recommended.	
	for cloning into TA vector.		
	Cautions for using Robust Buffer ( <i>Taq</i> )		
	Robust Buffer induces maximum enzymatic activity. To avoid production of undesirable		
	smear bands in gel electrophoresis analysis, the optimal reaction time is recommended as		
	follows: 1) about 5 to 10 seconds / kb elongation time for template up to 8 kb, and about 15		
	seconds / kb for up to 14 kb; 2) roughly the same elongation time is set with 2-step PCR (shuttle		
	PCR) and 3-step PCR; 3) extend the	elongation time by short steps when amplification is not	
	seen. Amplification can be detected	more rapidly by adopting 2-step PCR.	
Related product	02-001 Taq DNA Polymerase with Standard Buffer (+dNTPs)		
	02-021 Pfu DNA Polymerase with Standard Buffer (+dNTPs)		
Please note: All proc	ducts are FOR RESEARCH USE ONL	Y. NOT FOR USE IN DIAGNOSTIC	
PROCEDURES. NO	OT FOR MILITARY USE.		



Data Images: 02-002 Taq DNA Polymerase with Robust Buffer (+dNTPs)

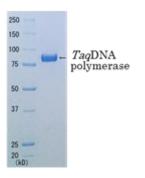


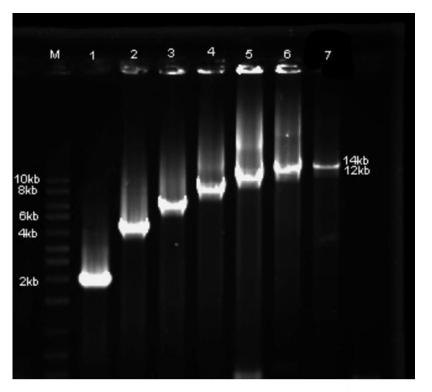
Fig.1 SDS-PAGE of TaqDNA polymerase

## Protocols for PCR

Examples of PCR conditions for the amplification of various sizes of  $\lambda$ DNA (results shown in Fig.2)

## 2 kb, 4 kb

94 °C 95 °C 65 °C	$ \begin{array}{c} 1 \text{ min} \\ 5 \text{ sec} \\ 20 \text{ sec} \end{array} \right\} 25 \text{ cycles} $		
6 kb			
94 °C 95 °C 65 °C	$\begin{array}{c} 1 \text{ min} \\ 5 \text{ sec} \\ 1 \text{ min} \end{array} \right\} 25 \text{ cycles}$		
8 kb			
94 °C 95 °C 65 °C	$ \begin{array}{c} 1 & \min \\ 5 & \sec \\ 1 & \min 20 & \sec \end{array} \right\} 25 \text{ cycles} $		
10 kb, 12 kb			
94 °C	1 min		
14 kb			
94 °C 98 °C 68 °C 72 °C	$ \begin{array}{c} 1 \text{ min} \\ 5 \text{ sec} \\ 4 \text{ min} \\ 4 \text{ min} \end{array} $ $ \begin{array}{c} 30 \text{ cycles} \\ 4 \text{ min} \end{array} $		



M: marker, lane1: 2kb, lane2: 4kb, lane3: 6kb, lane4: 8kb, lane5: 10kb, lane6: 12kb, lane7: 14kb,

## Fig.2 PCR products obtained by using Robust buffer (agarose gel electrophoresis)